Phytophthora ramorum

a guide for Oregon nurseries

J. Parke, J. Pscheidt, and R. Linderman

Figure 1.— Ramorum shoot dieback and leaf blight on Viburnum x 'Dawn'.

newly described funguslike organism named Phytophthora ramorum was discovered in 1993 to cause leaf blight, stem canker, and tip dieback on nursery-grown rhododendrons and viburnums in Germany and the Netherlands. At about the same time, many tanoaks (Lithocarpus densiflorus) and oaks (Quercus sp.) in the San Francisco Bay Area were dying from a new disease. The cause of this "sudden oak death" was also Phytophthora ramorum.

In 2001, the disease was detected in forest sites near Brookings, in southwest Oregon, on tanoak, Pacific rhododendron, and evergreen huckleberry. A 9-square-mile area was guarantined, and infected plant material was cut, piled, and burned in an attempt to eradicate the disease. Although a few new infested forest sites have been detected since then, they are all within the original quarantined area.

In May 2003, P. ramorum was discovered in a wholesale nursery in Clackamas County, Oregon, on Pieris, Rhododendron, and Viburnum. In June 2003, P. ramorum blight was reported on rhododendron and other hosts at a retail outlet in Washington that was affiliated with the Clackamas nursery. An international shipment of rhododendrons is

the suspected source of contamination for the Oregon and Washington nurseries. In a separate incident, in June 2003, infected camellias shipped from California were found in two Jackson County, Oregon, nurseries; they were destroyed. The same California nursery also had shipped camellias to two Portland-area garden centers. Approximately 300 of these plants were sold during January to July 2003. A public recall was initiated in summer 2003 to check plants and to remove and replace any infected plants found.

Further state and federal regulatory actions have been implemented to help prevent importing infested nursery stock; for more information, visit the Oregon Department of Agriculture Web site at http://oda.state.or.us/ plant/ppd/path/SOD/index.html. Pacific Northwest nursery growers need to be alert for inadvertent introductions of this pathogen, especially when importing from areas with known infestations.

December 2003 EM 8840 • \$2.00 Jennifer Parke and Jay Pscheidt, Department of Botany and Plant Pathology, Oregon State University; Robert Linderman, USDA-ARS Horticultural Crops Research Laboratory, Corvallis, OR.



An up-to-date list of hosts and associated species is online at http://www.aphis.usda.gov/ppq/ispm/sod/list.pdf

Figure 2.—Initial symptoms of ramorum leaf blight on *Viburnum plicatum* var. *tomentosum* 'Mariesii', showing necrotic leaf spots and shoot dieback.

Hosts

In the United States, the host list is growing; as of this writing, it includes 28 species in 12 plant families. Several more plant species are associated with *P. ramorum* but are not currently regulated, pending experimental confirmation.

Laboratory tests indicate that many more plant species, both wild and cultivated, are potentially susceptible to *P. ramorum*. In Europe, nursery hosts include rhododendron

cultivars, Viburnum spp., Pieris spp., Hamamelis virginiana, and Camellia spp. The disease has been reported from nurseries and gardens in Germany, the Netherlands, Belgium, France, Poland, Spain, Italy, Sweden, and the United Kingdom. P. ramorum also has been detected in a park on mature red oak specimen trees (Quercus rubra and Q. falcata) and on beech (Fagus sylvatica), horse chestnut (Aesculus hippocastanum), and seedlings of holm oak (Quercus ilex) in woodlands in the U.K.

Table 1. Nursery hosts reported in Oregon, plant part infected, and known symptoms of *Phytophthora ramorum*.

Plant species	Common name	Symptoms
Camellia japonica	Japanese camellia	Leaflesions
Camellia sasanqua	Sasanqua camellia	Leaflesions
Pieris floribunda x japonica	Pieris x 'Brouwer's Beauty'	Leaf lesions, shoot dieback
Pieris formosa x japonica	Pieris x 'Forest Flame'	Leaf lesions, shoot dieback
Pieris japonica	Japanese pieris	Leaf lesions, shoot dieback
Rhododendron hybrids	Rhododendron	Leaf lesions, shoot dieback; death of mature plants
Viburnum x bodnantense	Bodnant viburnum	Stem lesions
Viburnum plicatum var. tomentosum	Doublefile viburnum	Leaf lesions, shoot dieback; death of mature plants

Symptoms

Phytophthora ramorum causes different symptoms on different hosts. "Sudden oak death" on tree species is characterized by "bleeding" cankers that girdle the trunks of tanoaks (Figure 9, page 7) and some other oak species. On Rhododendron, Pieris, Viburnum, Camellia, and evergreen huckleberry, the disease is characterized by leaf blights and shoot diebacks and is more appropriately called ramorum leaf blight or ramorum shoot dieback. Symptoms on rhododendron (Figures 10–12, back page) may be indistinguishable from those caused by other Phytophthora species. The leaf petiole and midrib may be discolored, or the leaf tip or entire leaf blade may be necrotic (Figure 10). Leaf spots can occur where water accumulates on the leaf margins (Figure 11). Shoots die back when disease is severe (Figure 12). On Viburnum, infected leaves may die and fall off, leaving dark, leafless stems (Figures 1-3). In more severe infections, Viburnum can be killed. On Pieris, infected leaves turn a dark brown (Figure 7, page 5). Young shoots and leaves of Pieris are very susceptible to infection. Other hosts such as camellia may be infected but have only subtle symptoms, such as small leaf lesions on the lower leaves (Figures 5–6, page 4). Infected leaves on these hosts often fall off.

Biology

Phytophthora ramorum is a funguslike organism well adapted to the cool, wet conditions of the Pacific Northwest and at the same time tolerant of heat and drought. Unlike most *Phytophthora* species that

infect roots, *P. ramorum* is mainly a foliar pathogen. It produces several spore types, which helps the organism survive and spread (Figure 4). Spores landing on wet leaves or stems germinate and infect the plant. Young leaves are especially susceptible. Within a few days,

sporangia are produced, and they release tiny,



swimming spores (zoospores). The sporangia themselves can also detach, germinate, and infect. Sporangia and zoospores can be moved with windborne rain, in irrigation water, or with water splashed onto foliage. *P. ramorum* produces chlamydospores, which in other *Phytophthora* species are important for surviving unfavorable conditions. There are two mating types, designated A1 and A2.

The forest isolates from California and Oregon are the A2 type; the European isolates are mainly A1. Both types have been isolated from Oregon nurseries. If both mating types are in an infected plant, the pathogen could undergo sexual

Figure 3 (above).

—Viburnum
plicatum var.
tomentosum
'Mariesii' infected
with P. ramorum,
showing a
necrotic leaf as
well as defoliation
near the base of
the plant.

Figure 4 (at left).—Sporangia (clear) and chlamydospores (amber) of *P. ramorum* on an infected Oregon myrtlewood leaf.

Figure 5 (near right).— Symptoms on *Camellia* include leaf lesions and defoliation

Figure 6 (far right).—Leaf spots on *Camellia japonica* caused by *P. ramorum*.





Table 2. Woodland plant hosts, plant part infected, and known impact of *Phytophthora ramorum*.¹

Common name	State ²	Plant part infected and impact
Bigleaf maple	CA	Leaf lesions
California buckeye	CA	Leaf lesions
Madrone	CA	Leaf lesions; branch cankers
Manzanita	CA	Leaf lesions; dieback; stem and branch cankers
Toyon	CA	Leaf lesions; dieback; branch cankers
Tanoak	CA, OR	Bole cankers; leaf lesions; death of large trees
California honeysuckle	CA	Foliar lesions
Douglas-fir	CA	Foliar lesions; death of new shoots and small branches
Coast live oak	CA	Bole cankers; death of large trees
Canyon live oak	CA	Bole cankers; death of saplings; possible death of large trees
California black oak	CA	Bole cankers; death of large trees
Shreve oak	CA	Bole cankers; death of large trees
California coffeeberry	CA	Leaf lesions
Pacific rhododendron	CA, OR	Leaf lesions; shoot dieback; death of mature plants
Coast redwood	OR	Foliar lesions; branch cankers; death of sprouts; decline of saplings
Western starflower	CA	Leaflesions
Oregon myrtlewood, California bay laurel	CA, OR	Leaf lesions (tip necrosis or speckling
Evergreen huckleberry	CA, OR	Leaf lesions; shoot dieback
	Bigleaf maple California buckeye Madrone Manzanita Toyon Tanoak California honeysuckle Douglas-fir Coast live oak Canyon live oak California black oak Shreve oak California coffeeberry Pacific rhododendron Coast redwood Western starflower Oregon myrtlewood, California bay laurel	Bigleaf maple CA California buckeye CA Madrone CA Manzanita CA Toyon CA Tanoak CA, OR California honeysuckle CA Douglas-fir CA Cast live oak CA Canyon live oak CA California black oak CA Shreve oak CA California coffeeberry CA Pacific rhododendron CA, OR Western starflower CA Oregon myrtlewood, CAlifornia bay laurel

reproduction and form oospores. So far, oospores of *P. ramorum* have been observed only under laboratory conditions, so their role in disease epidemiology is not known. In other *Phytophthora* species, oospores are also thick-walled, like chlamydospores, and are important for surviving unfavorable conditions.

Disease prevention

Management efforts in Pacific Northwest nurseries are focused on eradicating the pathogen where it is found and preventing new infections. Early detection is vital to preventing disease spread. Practices useful in managing other foliar *Phytophthora* diseases also should help protect plants from infection by *P. ramorum*. The following strategies can reduce the risk of this disease in Pacific Northwest nurseries.

Exclusion and avoidance

- ▲ If importing nursery stock of trees or shrubs from any source (out of state or international), you must notify the Nursery & Christmas Tree Program Supervisor at the Oregon Department of Agriculture by fax (503-986-4786) or e-mail (quarantine@ oda.state.or.us). Plants must be certified as being free from *P. ramorum*. For more information on Oregon's import regulations, go to http://oda.state.or.us/plant/ppd/path/SOD/index.html Illegal importations may result in significant fines. The USDA's quarantine regulations regarding *P. ramorum* are online at http://www.aphis.usda.gov/ppq/ispm/sod
- ▲ Keep imported known hosts and plants associated with *P. ramorum* in a separate block at least 7 feet from other plants in the nursery for 3 to 6 months, to see the development of any symptoms that initially might have been masked by fungicides or delayed by weather.
- ▲ Do not transport oak firewood or other potentially infected plant materials from infested areas in California to Oregon.
- ▲ If you visit infested areas, wash your vehicle and shoes before traveling to disease-free areas.

Cultural management

- ▲ Familiarize yourself and your staff with the range of symptoms of *Phytophthora ramorum*. Check your plants often. Diseases caused by other *Phytophthora* species can cause similar symptoms. If you suspect *P. ramorum*, call the Oregon Department of Agriculture's Invasive Species Hotline, 1-866-INVADER, or your Nursery Inspector at 503-986-4644. Diagnosis, using several techniques, may take 1 to 2 weeks.
- ▲ While waiting for the diagnosis, do not move or ship symptomatic plants **or** any nearby plants; even if they look healthy, they may be contaminated.
- ▲ Be alert for symptoms on **any** shrub and tree species, not just those on the list of hosts and plant species associated with *P. ramorum*.
- ▲ Disinfect tools and shoes that may have been in contact with contaminated plants or potting media. Incinerate contaminated pots and trays or treat them with aerated steam to kill the pathogen.
- ▲ Propagate cuttings only from plants known to have been free of disease for several months. Sanitize cuttings to eliminate the pathogen; soak cuttings in a disinfectant before storage and/or sticking in rooting medium. Use clean, pathogen-free potting media and clean, new pots.

Figure 7.—Leaf and stem necrosis and shoot dieback on *Pieris japonica*.



- Manage irrigation to reduce the length of time that foliage is wet. If possible, increase watering intervals. Improve drainage to avoid puddling and splashing. Place pots on fast-draining surfaces.
- ▲ Use only clean water for irrigation.

 Treat irrigation water to kill spores of
 P. ramorum and other Phytophthora species.

 Remove and destroy any fallen leaf material, dead branches, or plants.

Table 3. Plant species associated with *P. ramorum* but not currently regulated, pending experimental confirmation.

Plant species	Common name	Location	
Abies grandis	Grand fir	CA	
Aesculus hippocastanum	Horse chestnut	E	
Arbutus unedo	Strawberry tree	E	
Camellia reticulata	Camellia	E	
Camellia× williamsii	Camellia hybrid	E	
Corylus cornuta	California hazelnut	CA	
Fagus sylvatica	Beech	E	
Kalmia latifolia	Mountain laurel	E	
Leucothoe fontanesiana	Drooping leucothoe	E	
Pieris formosa var. forestii	Chinese pieris	E	
Pieris formosa var. forestii × P. japonica	Pieris hybrid	E	
Pittosporum undulatum	Victorian box	CA	
Quercus falcata	Southern red oak	E	
Quercus ilex	Holm oak	E	
Quercus rubra	Northern red oak	E	
Rhamnus purshiana	Cascara	OR	
Rubus spectabilis	Salmonberry	OR	
Syringa vulgaris	Lilac	E	
Taxus baccata	European yew	E	
Toxicodendron diversiloba	Poison-oak	OR	
Vaccinium vitis-idaea	Lingonberry	E	
Viburnum davidii	David viburnum	E	
Viburnum farreri (=V. fragrans)	Fragrant viburnum	Е	
Viburnum lantana	Wayfaringtree viburnum	E	
Viburnum opulus	European cranberry bush viburnum	Е	
Viburnum x burkwoodii	Burkwood viburnum	E	
Viburnum carlcephalum			
x V. utile	Viburnum hybrid	E	
Viburnum × pragense	Prague viburnum	Е	

¹CA=California, E=Europe, OR=Oregon

Protection and suppression with fungicides

Use fungicides as preventive treatments on known host plant species. Most fungicides used to manage *Phytophthora* do not kill this organism. They can only prevent the organism from becoming established. They also can prevent continued growth if the organism is already inside the plant—thereby masking symptoms that might have developed. Once chemical activity has subsided with time (about 3 to 6 months), the organism can resume growth within infected plants. This is why plants from other nurseries must be held for several months to see whether symptoms become evident.

If you choose to use fungicides, alternate among ones from different fungicide families with different modes of action. For specific fungicide recommendations, consult the *PNW Plant Disease Management Handbook* or An Online Guide to Plant Disease Control at http://plant-disease.ippc.orst.edu/index.cfm

Detection and eradication

If *P. ramorum* is found in your nursery, ODA and USDA will work with you, at no charge, to prevent further disease spread, to eradicate the infestation, and to monitor your nursery to verify when it is free from *P. ramorum*. USDA is drafting uniform procedures for dealing with nursery infestations. Check the regulatory action plan on the ODA and USDA-APHIS Web sites for up-to-date information. The plan may include many of the following provisions.

- ▲ Samples will be taken from symptomatic plants to confirm the pathogen's identity. Testing usually will be part of the annual survey.
- ▲ Samples will be taken from other plants in the nursery to determine the extent of the infestation.
- ▲ Host plants will not be sold or moved during the investigation (generally, less than 2 weeks).
- ▲ Blocks of plants containing infested plants will be destroyed by deep burial or by incineration under ODA's direct supervision.

- ▲ Healthy host plants within 10 meters of infested blocks will be held for 90 days* to observe symptoms. These plants may not be treated with fungicides during this time so that symptoms will not be masked.
- ▲ Nonhost plants can be shipped during this time.
- ▲ Surrounding properties will be surveyed for *P. ramorum*.
- ▲ Trace-back investigations will be made to determine the source of infected plants, and trace-forward investigations will be made if any plants from the infested block(s) were shipped.
- ▲ ODA will monitor water, potting mix, soil, and plants for the presence of *P. ramorum*. When tests have been negative for 90 days*, the nursery may once again ship host plants.
- ▲ The nursery will be tested each spring for the next 2 years as a precaution.



Oregon State University Extension Service.
An Online Guide to Plant Disease Control.

http://plant-disease.ippc.orst.edu/
index.cfm

Pscheidt, J.W. and C.M. Ocamb, eds. Pacific Northwest Plant Disease Management Handbook (revised annually). Corvallis, OR: Extension Services of Oregon State University, University of Idaho, and Washington State University.

California Oak Mortality Task Force
http://suddenoakdeath.org/

Davidson, J.M., S. Werres, M. Garbelotto, E. Hansen, and D.M. Rizzo. 2003. Sudden Oak Death and associated diseases caused by *Phytophthora ramorum*. Plant Health Progress, Plant Management Network International.

▲ http:/www.plantmanagementnetwork. org/pub/php/diagnosticguide/2003/sod/





Oregon Department of Agriculture, Sudden Oak Death Alert.

▲ http://oda.state.or.us/plant/ppd/path/ SOD/index.html

USDA Animal and Plant Health Inspection Service (APHIS), Pest Detection and Management Programs, Invasive Species and Pest Management: Sudden Oak Death.

▲ http://www.aphis.usda.gov/ppq/ispm/sod/

Figure 8 (above left).—Foliar symptoms on susceptible oaks include a darkened petiole and midrib, as on this tanoak leaf, or necrosis at the leaf base.

Figure 9 (above right).—Bleeding canker on the trunk of a mature tanoak.

^{*}The 90-day period must be during active plant and pathogen growth; otherwise, the waiting period may be extended.





Acknowledgments

In the preparation and printing of this report, the authors gratefully acknowledge the technical assistance and financial support of the Oregon Association of Nurseries, a 1,600-member trade association representing the interests of those who grow, handle, and retail ornamental horticultural plant material.

OREGON ASSOCIATION OF **NURSERIES** Photos Figure 1, Jan Hedberg, ODA;

Figure 10 (above plants with ramorum leaf blight. Plant in was killed by ramorum leaf background shows early symptoms of ramorum leaf blight on lower

Figure 11 (above

Figure 12 (at right, below).— Ramorum shoot dieback on wild



Figures 5 and 6, Cheryl Blomquist, California Department of Food and Agriculture; Figure 7, Nancy Osterbauer, ODA; Figures 9 and 12, Everett Hansen, Oregon State University. All other figures by Jennifer Parke.

© 2003 Oregon State University. Produced and distributed in furtherance of the Acts of Congress of May 8 and June 30, 1914. Extension work is a cooperative program of Oregon State University, the U.S. Department of Agriculture, and Oregon counties. Oregon State University Extension Service offers educational programs, activities, and materials—without discrimination based on race, color, religion, sex, sexual orientation, national origin, age, marital status, disability, and disabled veteran or Vietnam-era veteran status. Oregon State University Extension Service is an Equal Opportunity Employer.

Published December 2003.